

Updated cultivation methods for *Palmaria palmata*

– *new insights towards a reliable production*

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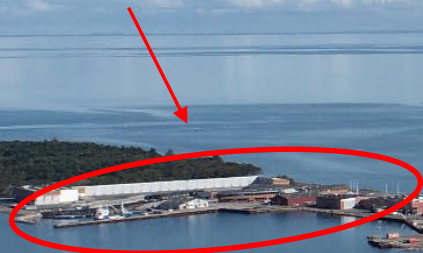
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Section for Coastal Ecology



➤ MSc. 2013 (AU)
Kelp cultivation & nutrient uptake and salinity

➤ Kelp farmer 2013-2015

➤ PhD. 2020
Palmaria palmata

- Background
- Life cycle
- Optimizing hatchery



DTU Aqua
National Institute of Aquatic Resources



Investigating hatchery and cultivation methods
for improved cultivation of
Palmaria palmata

Peter Søndergaard Schmedes

PhD thesis, April 2020



Palmaria palmata

- High reputation - dried snack / unique taste
- Valuable contents - new applications
- Demands a scalable cultivation practice and optimized methods



Credit: Werner & Dring 2011



Cultivation workflow



Hatchery seeding method based on gravity - works, but is in-efficient



Bottlenecks

- High requirement of spore donors for substrate seeding
- Only one seeding method with short window
- Variable seeding quality and loss of spores

Requirement of spore donor tissue (kg/t FW harvest)

~ 72 kg FW/t FW harvest

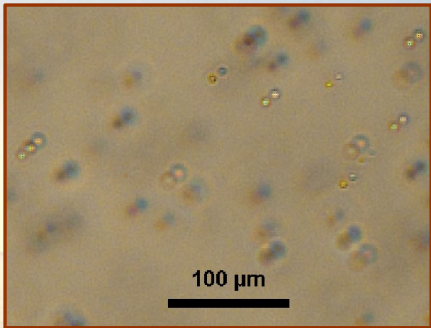
< 1 kg FW/t FW harvest

Life cycle of *P. palmata*

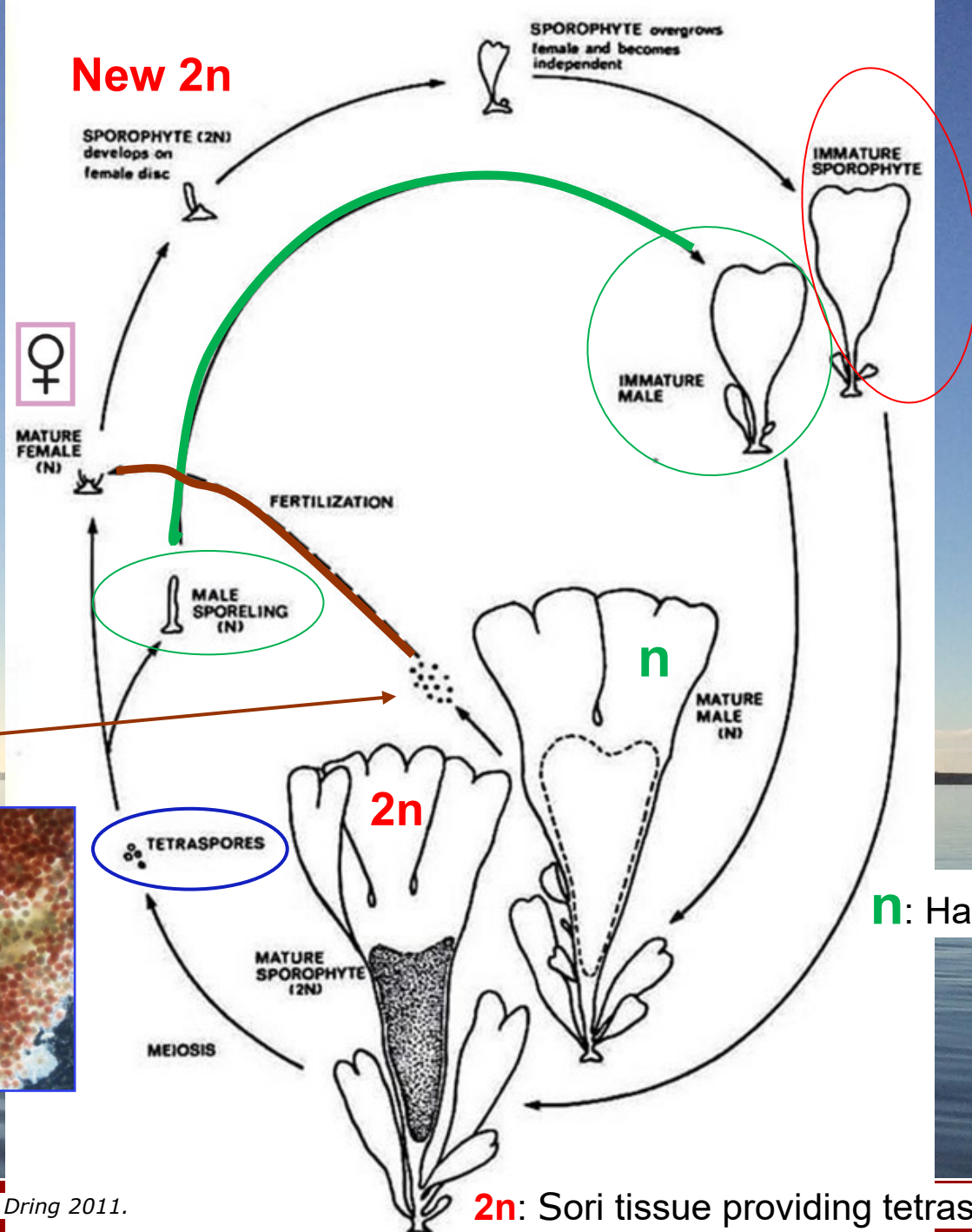
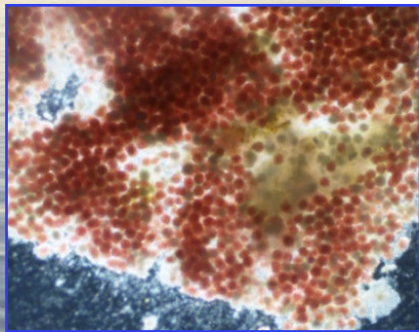
□ Basics for hatchery improvement

In the hatchery

- Spore dispersal vs. aggregates
- Fertilization improves spore use



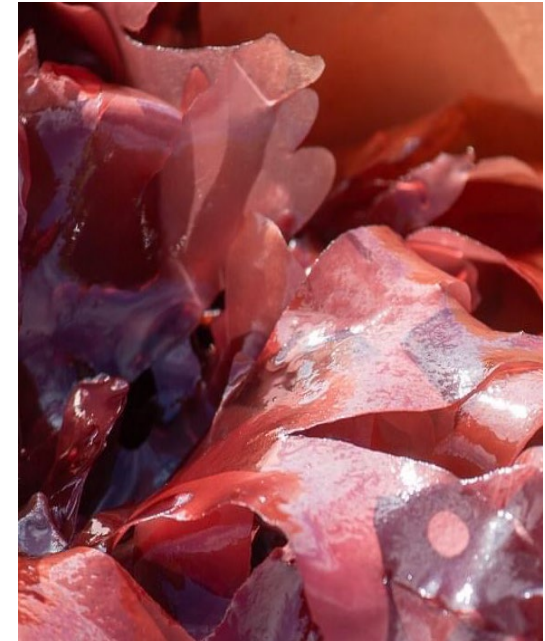
Male gametes in sea water



n: Haploid male gametophyte

2n: Sori tissue providing tetraspores

Backwards review of challenges related to the cultivation work flow



✓ Harvest

Sufficient harvest yield of clean biomass

Backwards review of challenges related to the cultivation work flow



✓ **Cultivation**

- Deployment unit (rope, net, sheet)
- High specific biomass yield
- Ease of handling (hatchery, deployment, harvest)

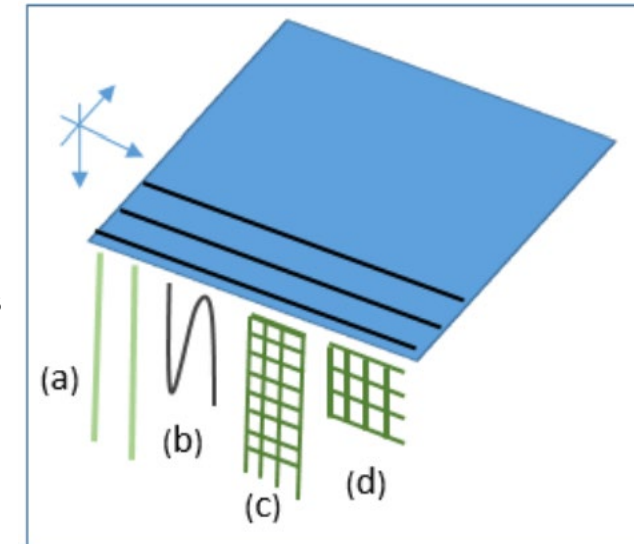
✓ **Harvest**

- Sufficient harvest yield of clean biomass

Longline system

Deployments:

- (a) Single droppers
- (b) Loops
- (c) Vertical net
- (d) Horizontal net



Backwards review of challenges related to the cultivation work flow



? Nursery ?

Optimal density of seedlings ($10-50 \text{ cm}^{-1}$) ~ seeding density ($20-100 \text{ spores cm}^{-1}$)
 Seedling size (0.5-4 cm) ~ duration

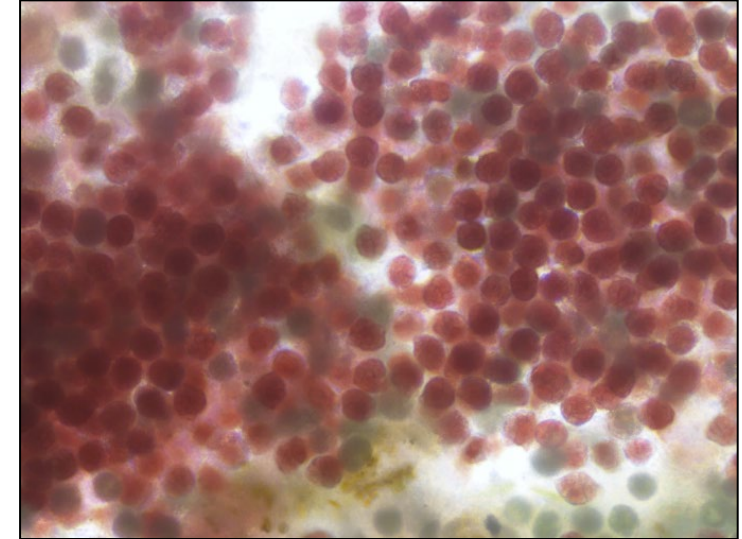
✓ Cultivation

Deployment unit (rope, net, sheet)
 High specific biomass yield
 Ease of handling (hatchery, deployment, harvest)

✓ Harvest

Sufficient harvest yield of clean biomass

Backwards review of challenges related to the cultivation work flow



? Hatchery seeding ?

Timing of spore release and spore yield ~ duration ?

Even spore settlement ~ seeding method (sori vs propagules) ?

? Nursery ?

Optimal density of seedlings ($10-50 \text{ cm}^{-1}$) ~ seeding density ($20-100 \text{ spores cm}^{-1}$)

Seedling size (0.5-4 cm) ~ duration

✓ Cultivation

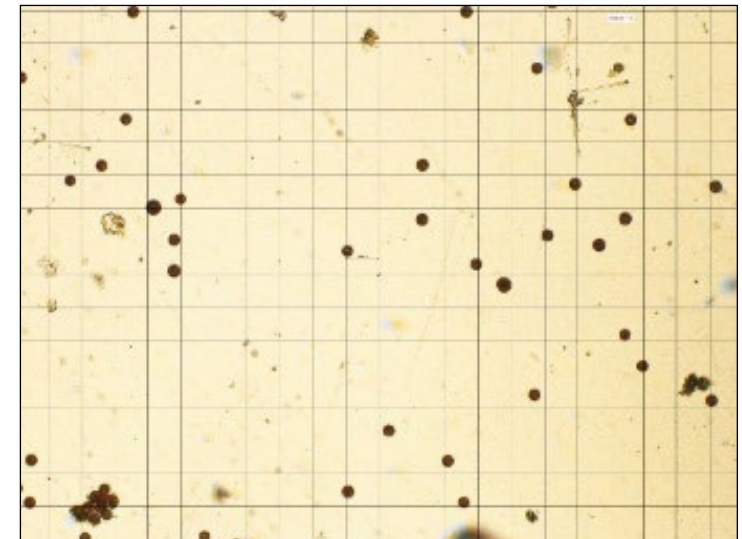
Deployment unit (rope, net, sheet)

High specific biomass yield

Ease of handling (hatchery, deployment, harvest)

✓ Harvest

Sufficient harvest yield of clean biomass



Backwards review of challenges related to the cultivation work flow



? Collection of spore donors ?

Timing of season, sporophyte recognition?

? Hatchery seeding ?

Timing of spore release and spore yield ~ duration ?

Even spore settlement ~ seeding method (sori vs propagules) ?

? Nursery ?

Optimal density of seedlings ($10-50 \text{ cm}^{-1}$) ~ seeding density ($20-100 \text{ spores cm}^{-1}$)

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✓ Cultivation

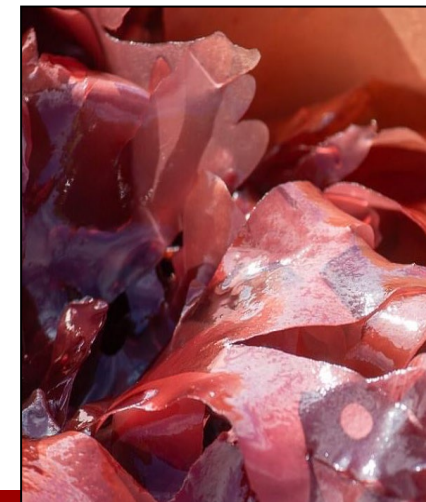
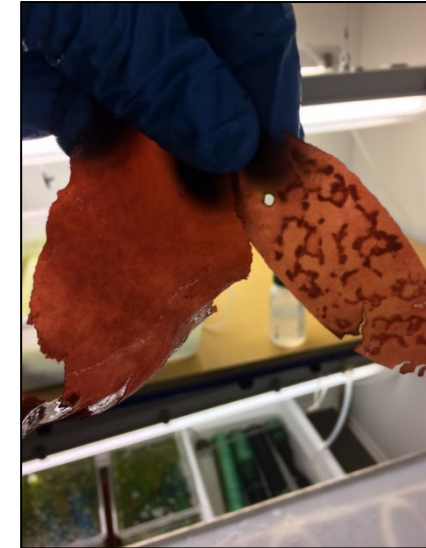
Deployment unit (rope, net, sheet)

High specific biomass yield

Ease of handling (hatchery, deployment, harvest)

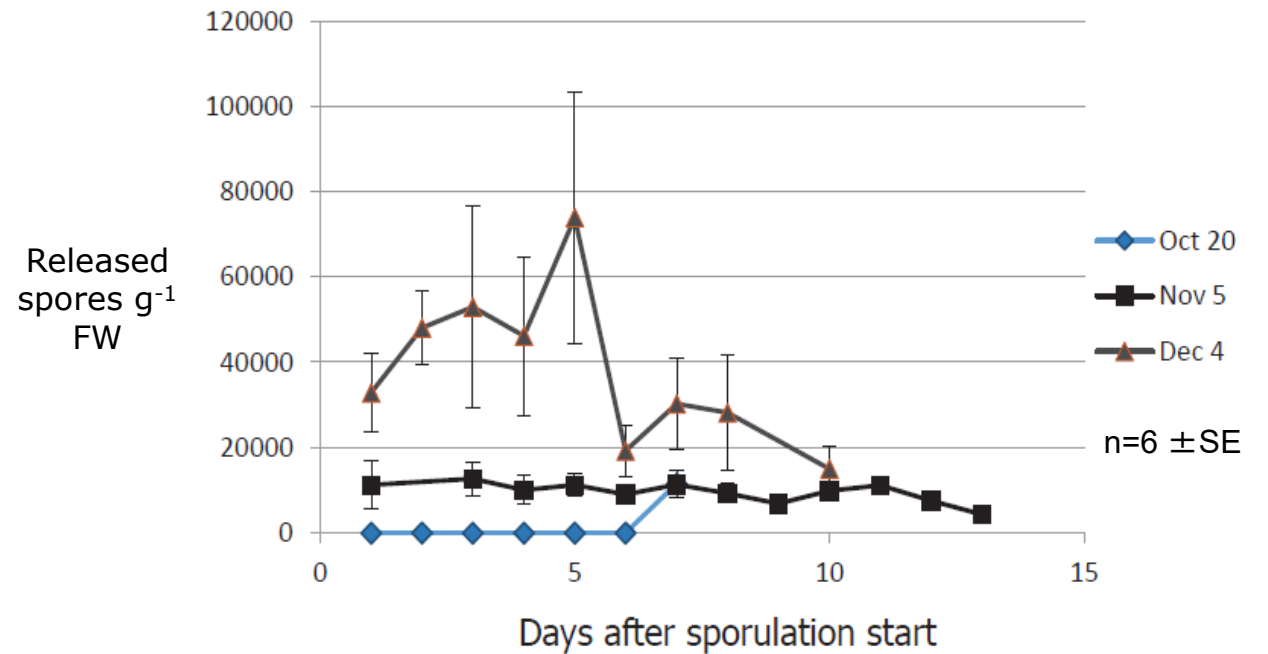
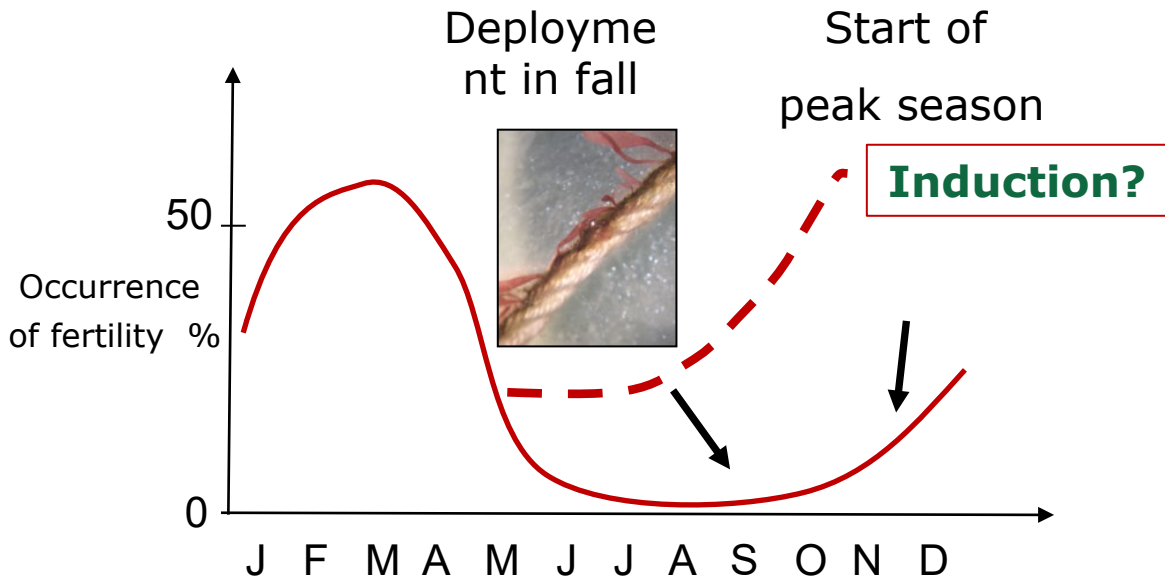
✓ Harvest

Sufficient harvest yield of clean biomass



Seasonality in reproduction

How to secure access to spore donors?

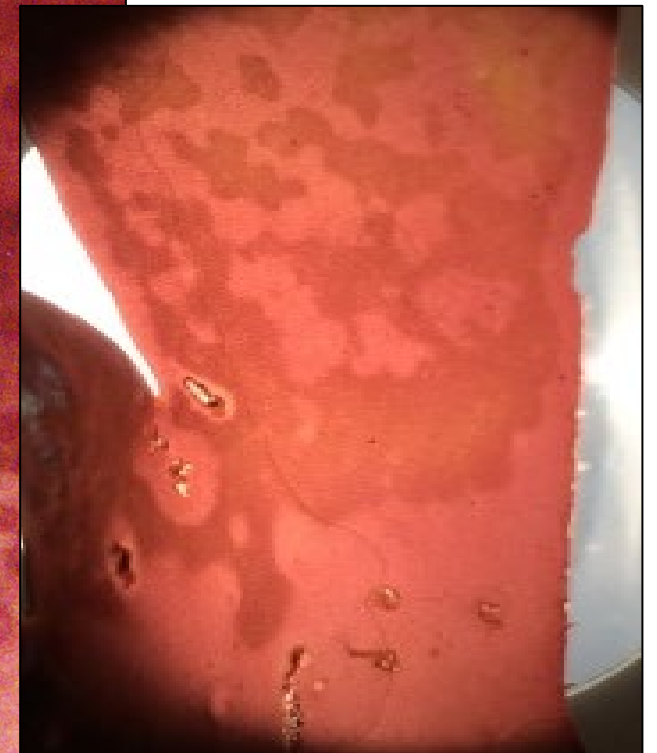
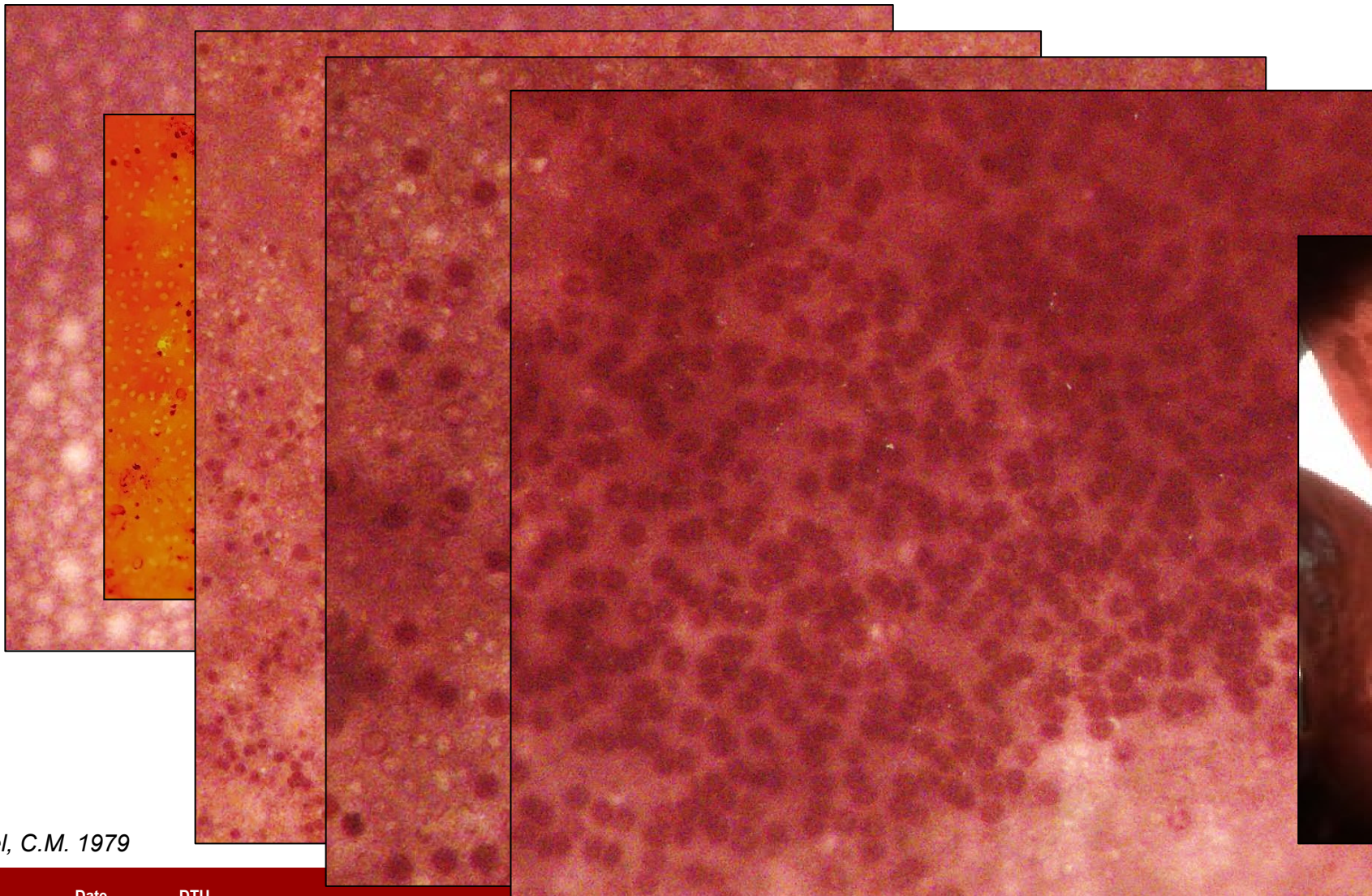


- Germination and development of seedlings require 1-3 months to reach a size min. 5 mm
- **Manipulating env. parameters – but the response time is still long – induction not yet effective strategy!**
- **Accelerate/pause sporangia maturation**

Identifying sporophyte and male gametophyte Sporangial initials in the cortex layer



2n or n?



Pueschel, C.M. 1979

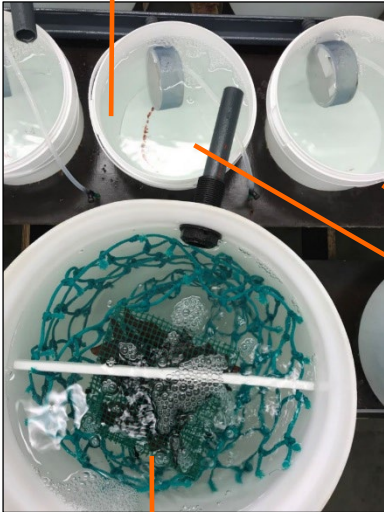
Focus on improving spore use efficiency in hatchery seeding

- ✓ Less spore donors
- ✓ Better dispersal
- ✓ Improve survival

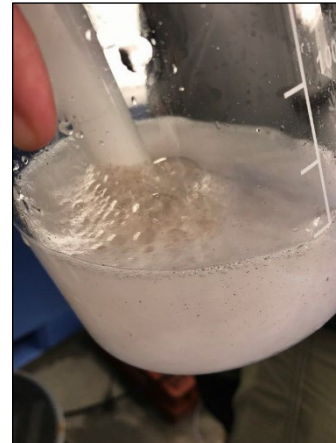
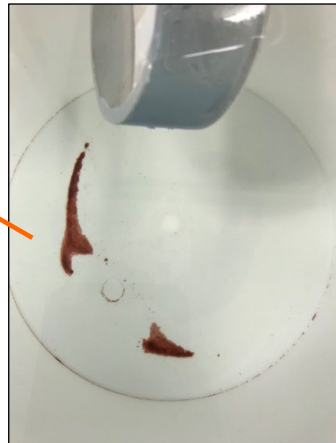


Tumble flow-through

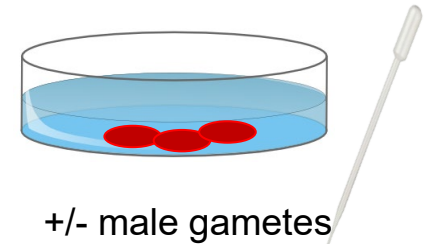
Retain "effluent spores"



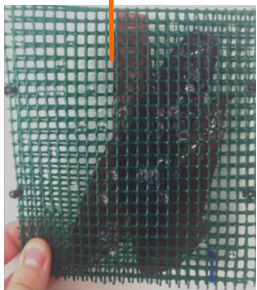
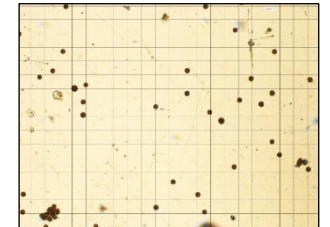
- Count spores + seedlings (day 3, 19, 32)
- Net seeding efficiency = $\frac{\text{spores}_{\text{NET}}}{(\text{spores}_{\text{NET}} + \text{EFFLUENT})}$



Include fertilization

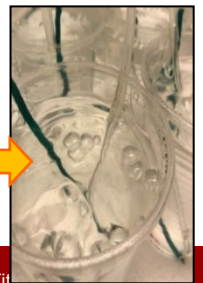
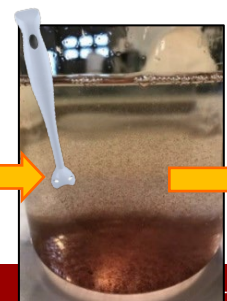
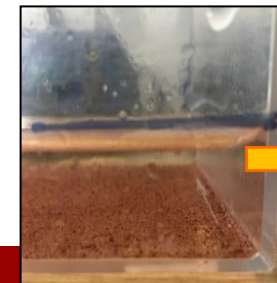


+/- male gametes



- Packages with increasing amount of spore donors i.e., 5, 10, 15 gFW
- Re-use: Seeded 3 nets during 9 days (3 days each)

Germination Maceration Agitation



Results on hatchery seeding

Flow-through seeding

- Net seeding efficiency = 16 %
- Even spore settlement (day 3)
- 9 seedling cm^{-1} after 32 days



- Nursery duration of 1 vs. 9 months

GMA-seeding ropes

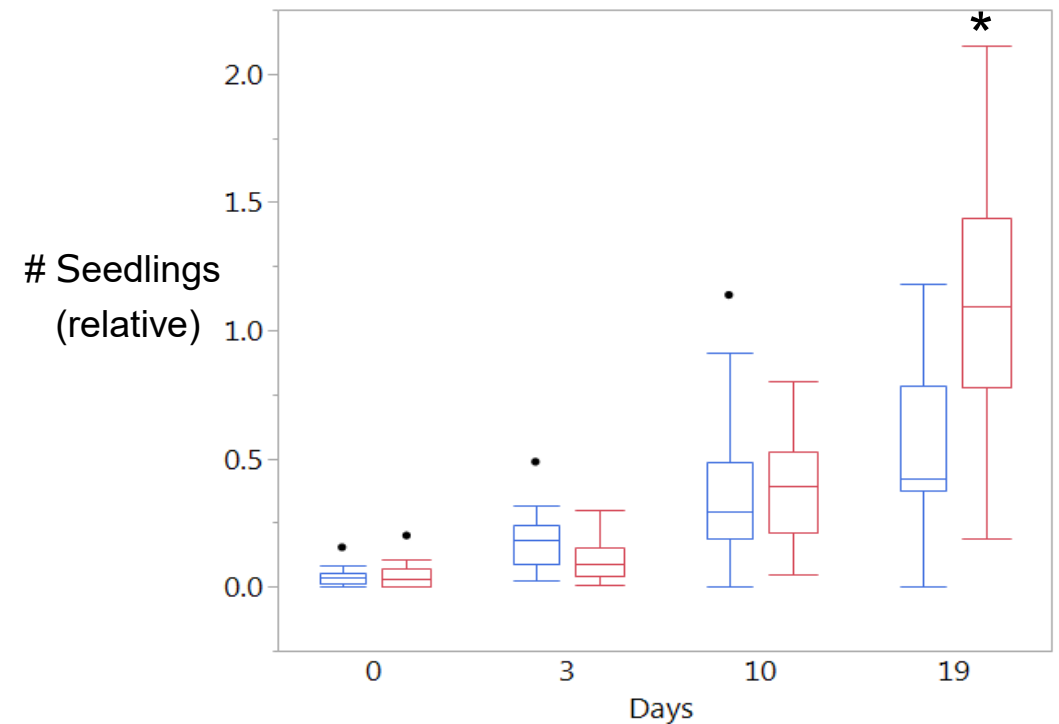
- Successful re-attachment for germlings detached at day 29 and 39 (not day 240)
- 60 seedlings cm^{-1} (high concentration used)



- Nursery duration of 9 months (density lowered)

Fertilization step

- Higher seedling density on day 20 (activation of females)



(Schmedes et al., 2019, Alg Res; Schmedes & Nielsen, JAPH, 2019)

Biomass yields from a cultivation trial

9 months nursery



1. Harvest 2020 (cropping)



Over-summering



2. Harvest 2021 (cropping)



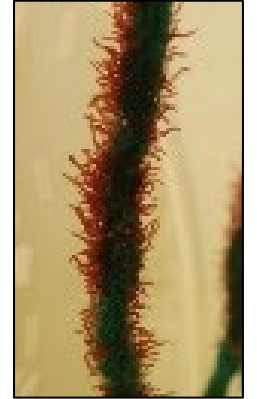
Substrate	Harvest year	Cultivation depth (m)	Avr. biomass yield kgFW/m longline	Max. yield kgFW/m longline
Net	2020	1.5	2.9 kg (n=3)	4.9 kg (n=1)
GMA-rope	2020	1.5	1.6 kg (n=3)	1.7 kg
Net	2021	1.5	2.4 kg (n=3)	2.8 kg
Total (nets)			5.3 kg (n=3)	7.7 kg (n=1)

Perspectives for upscaling hatchery production

- ❑ Several ways to improve hatchery production of *Palmaria palmata*
 - Increase hatchery seeding efficiency (m substrate / g spore donor tissue) ~ minimum or re-use
 - Optimize spore settlement distribution ~ substrate feature & spore dispersal
 - Maximize spore survival ~ activate female gametophytes & water quality & alternative seeding (GMA)

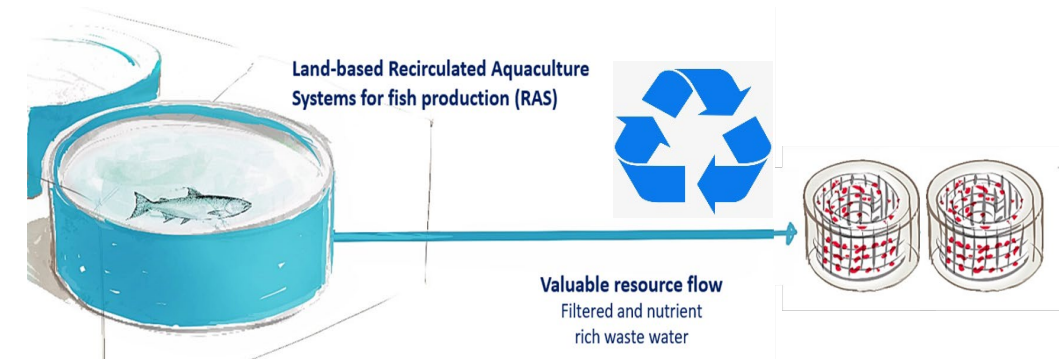
- ❑ Compare seeding efficiency of two different seeding methods:
 - The horizontal gravity-based (Dring & Werner 2011)
 - 23g/m substrate (1.8 kg FW fertile sporophytes to seed a net equivalent to 82 m rope)
 - 136 kg FW fertile sporophytes to seed enough nets for 100 m longline deployment

 - Vertical flow-through seeding system (Schmedes 2020)
 - 0.1-0.3 g/m substrate (5-15 g FW fertile sporophytes to seed a net equivalent to 46 m rope)
 - 0.6-2 kg FW fertile sporophytes to seed enough nets for 100 m longline deployment
 - **Improvement of factor 68-224 in hatchery seeding efficiency**



Landbased palmaria cultivation - why?

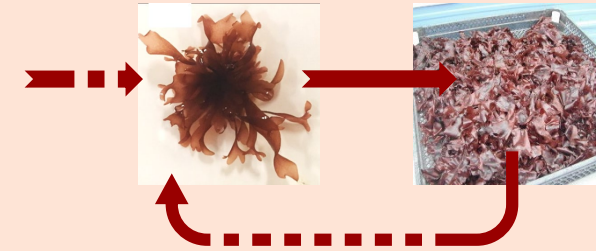
- It is worth the effort – high market prices
 - One of the most valued seaweed species for food
 - » Prices up to **250 € per kg** (dry)
- Provides extra possibilities
 - Higher control → **high quality**
 - Integrated **multi-species culture** practise
 - » Nutrient removal/nutrient re-use
 - » Green branding





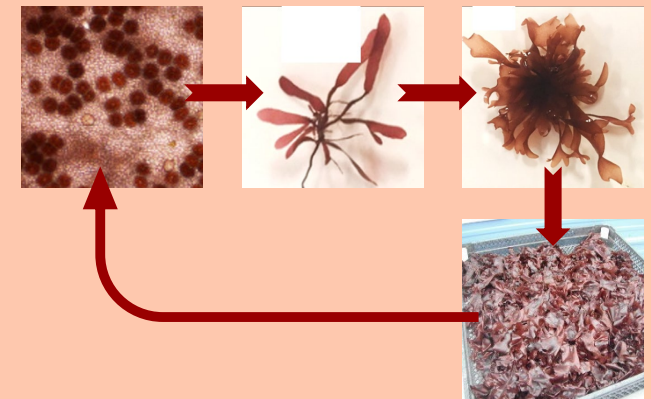
Propagation of **wild** harvested biomass

- Vegetative growth
- Easy to operate
- Starter culture from wild resources
 - Frequent (pr. season)
 - Renewal



Spore-based propagation

- DK: Lack of hard substrate
Wild beds are difficult to access
- Sustainability (less impact on wild stocks)
- Starter culture from wild
 - Potentially independent in future





Germinate

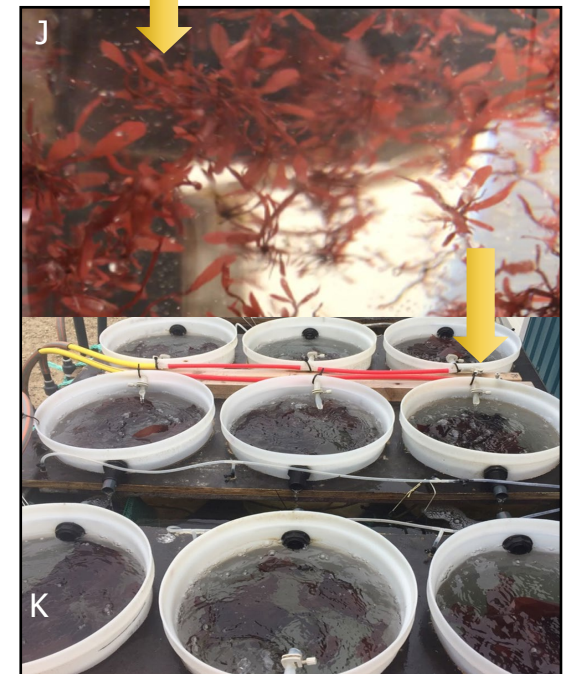
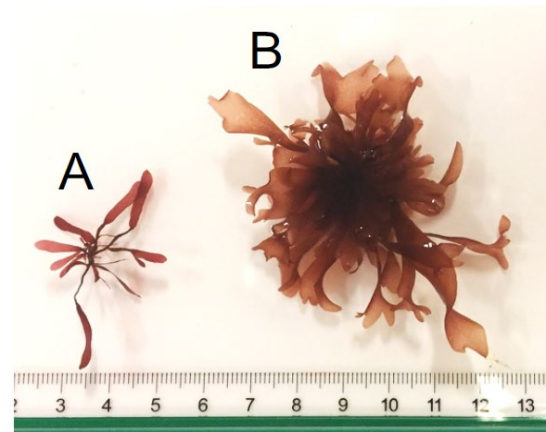
Macerate

Re-attach



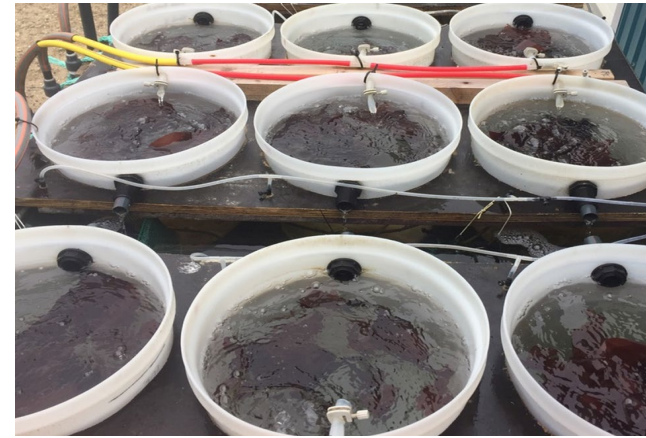
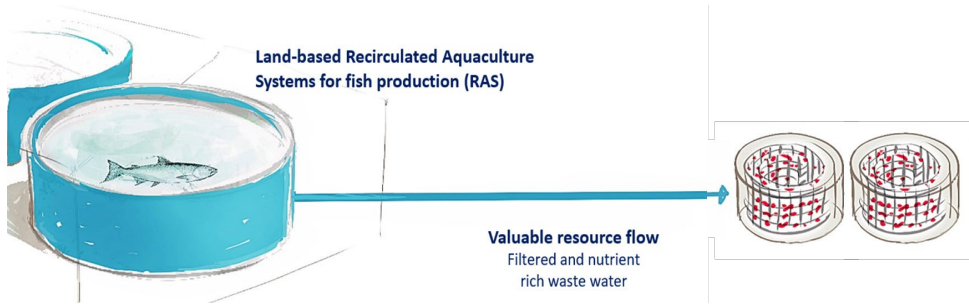
Seedling bank:

Possible to keep at "skeleton scale" for minimum 3 years at low maintenance

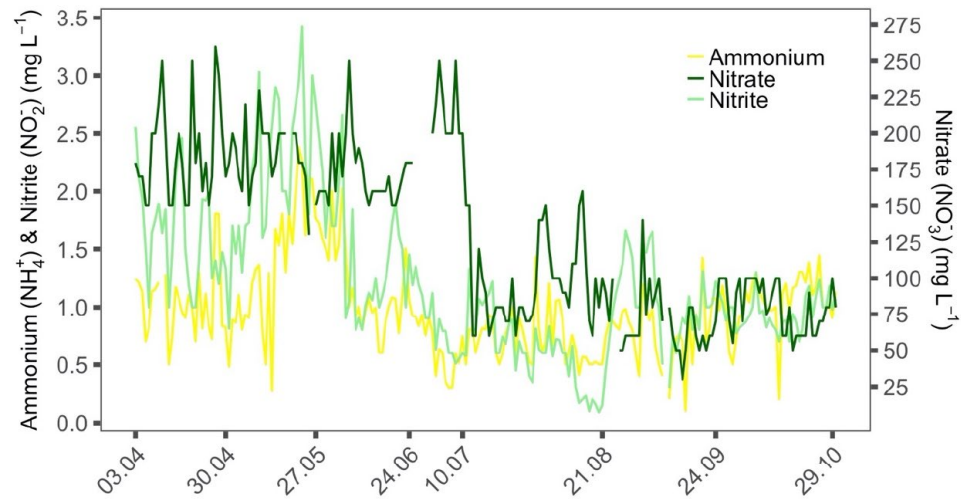




Master project: Jørgen Levinsen
 Cultivation of *Palmaria palmata* in
 land-based IMTA systems

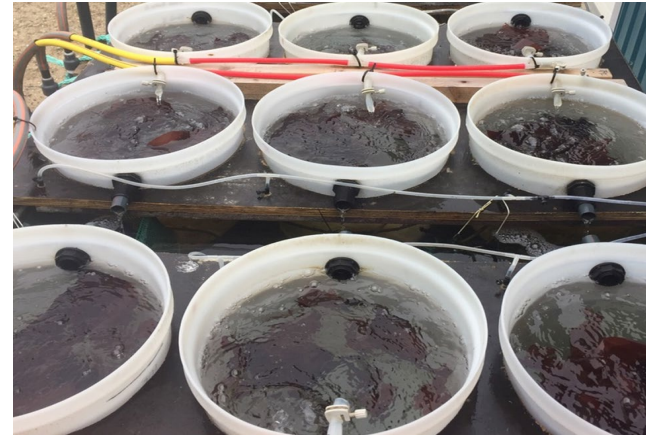
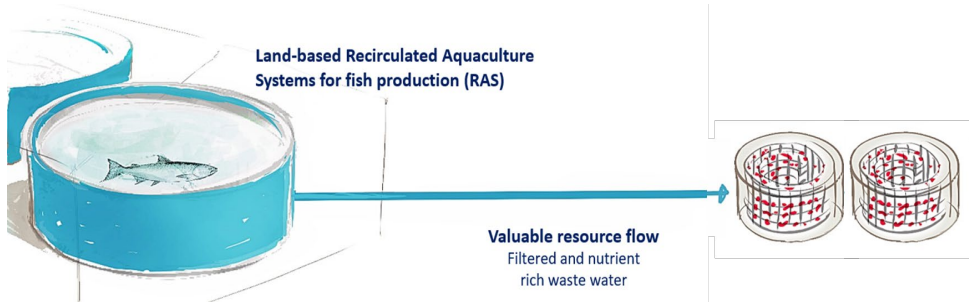


Flow through bubbling tanks (30 l) with
 process water from landbased salmon farm.

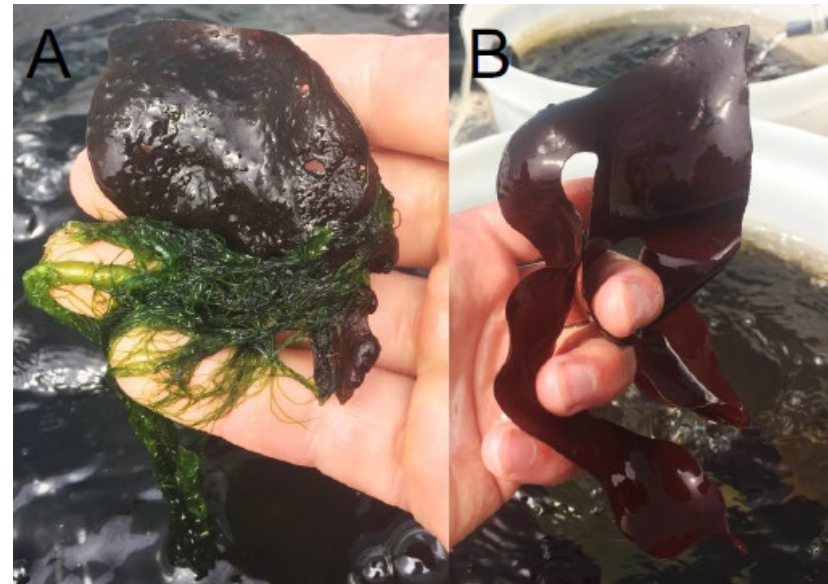
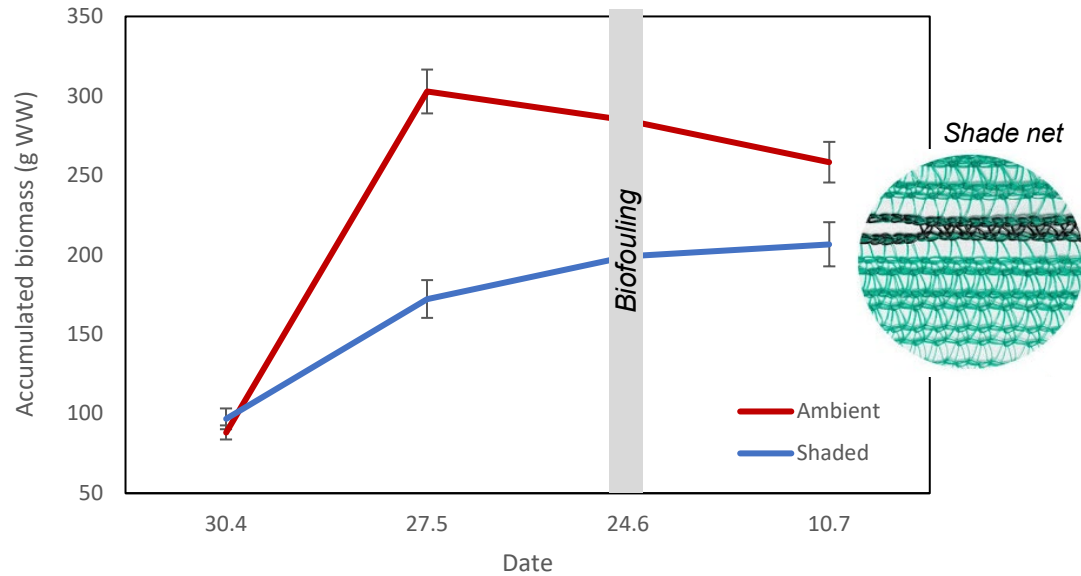




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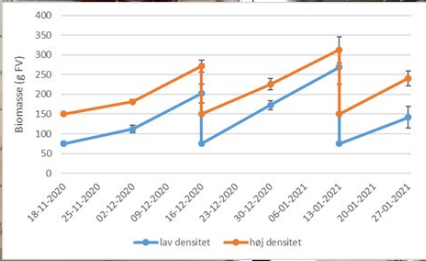
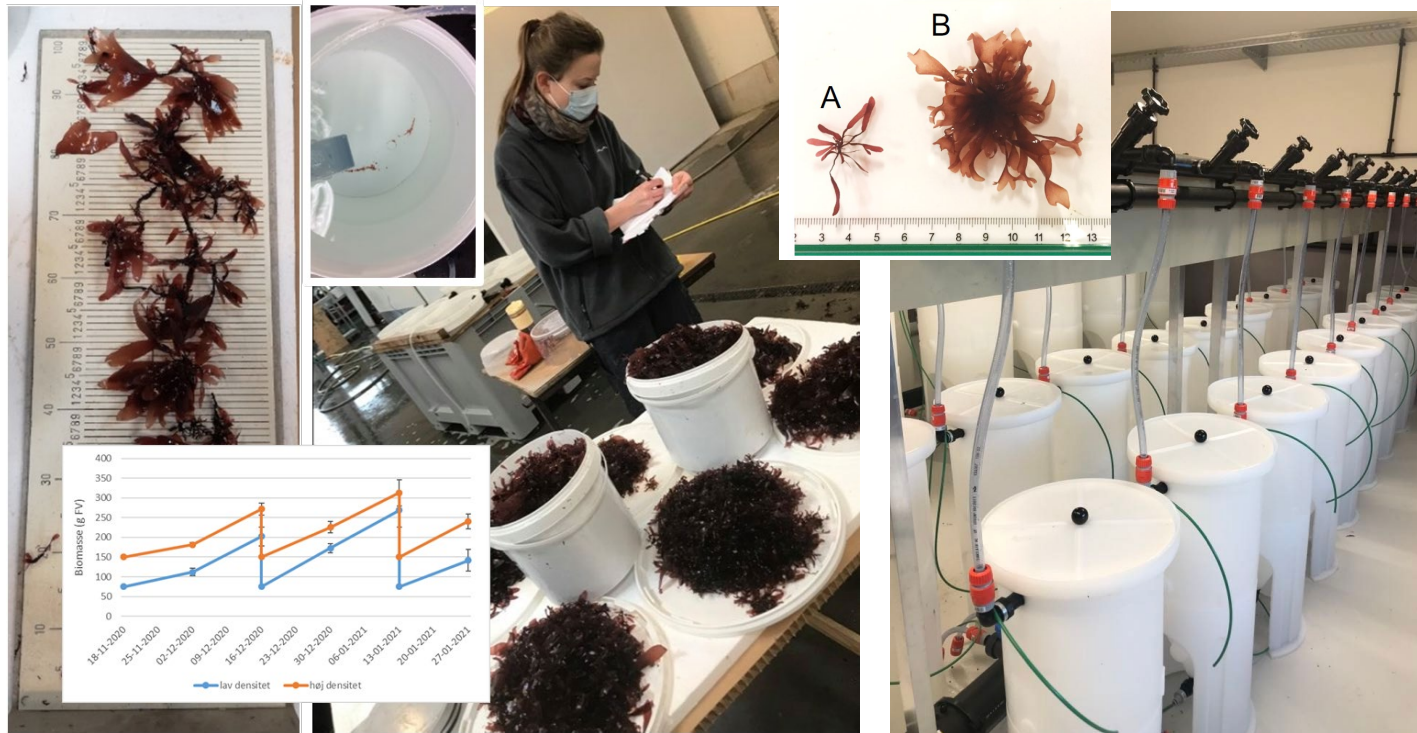


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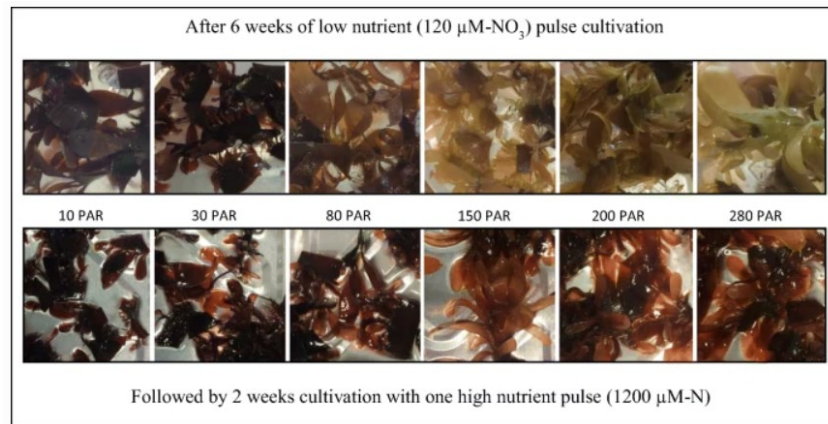


Ambient

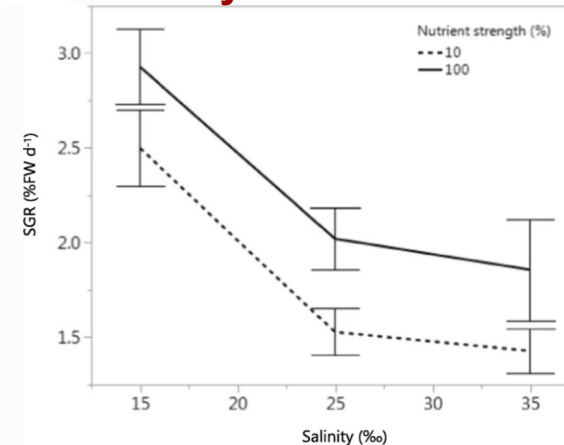
Shaded



Nutrient pulse



Salinity



Current research:

- Light (e.g. red light to reduce fouling)
- Nutrients (e.g. pulse)
- Density
- Salinity (e.g. low/fluctuating salinity tolerance)
- Maintenance (man power need)
- Fouling control (e.g. chemical, "environmental" triggers)

Journal of Applied Phycology (2020) 32:4099–4111
<https://doi.org/10.1007/s10811-020-02248-4>

Productivity and growth rate in *Palmaria palmata* affected by salinity, irradiance, and nutrient availability—the use of nutrient pulses and interventional cultivation

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Received: 16 January 2020 / Revised and accepted: 9 September 2020 / Published online: 13 October 2020
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Abstract
 Land-based cultivation of the rhodophyte *Palmaria palmata* is promising for high productivity and nutrient mitigation, yet the cultivation strategy and the knowledge of the effect of various environmental factors are incomplete. In a two-phased cultivation trial, marginal proliferations were used as seedstock to test the impact of irradiance (10–280 µmol photons m⁻² s⁻¹ photosynthetically active radiation; PAR) using sequential nutrient phases of pulse additions (10% vs. 100% F/2+) on specific growth rate (SGR) and productivity (exp. 1). The effect of salinity (15–35‰) and nutrient concentration (10 vs. 100% F/2+) on frond growth was investigated (exp. 2). The SGR peaked at 200 µmol photons m⁻² s⁻¹ PAR in both nutrient phases with mean SGR of 6.86 ± 0.4% day⁻¹ (mean ± SE, n = 3). Above 80 µmol photons m⁻² s⁻¹ PAR, thalli turned pale green after 3 weeks at low nutrient. Shifting to a high nutrient cultivation, thalli recovered their red color after 10 days, even at 280 µmol photons m⁻² s⁻¹ PAR and significantly upshifted SGR, dry matter (DM), nitrogen (N), phosphorus (P), and ash content by 79.3, 56.0, 27.3, and 16.4%, respectively. Peak productivity in DM (1.17 g DM m⁻² day⁻¹), carbon (C) (406.41 mg C m⁻² day⁻¹), N (20.61 mg N m⁻² day⁻¹), and P (2.06 mg P m⁻² day⁻¹) coincided with SGR. Salinity significantly affected SGR of *P. palmata* and peaked at 15‰. This study highlights the use of marginal proliferations seedstock, nutrient pulses, and intervention practice for biomass propagation of *P. palmata* while avoiding epiphytes to boost N removal.

Keywords Rhodophyta · Seedstock · RAS seaweed · Nutrient pulses · N-starvation · N-removal · Bracketish salinity

Introduction
 The rhodophyte seaweed *Palmaria palmata* (L.) F. Weber & D. Mohr is highly valued as nutritious snack food, known as dulse, and as a promising source of bioactive compounds for aquaculture feed, cosmetics, and nutraceuticals (Holdt and Kraan 2011; Mouritsen et al. 2013; Moroney et al. 2015; Lopes et al. 2019). Currently, the supply of *P. palmata* for commercial products is sourced by hand harvesting wild populations, which is time-consuming and raises increasing concerns on overexploitation of wild seaweed populations (Ugarte and Sharp 2001; Monagari et al. 2017). Increasing demand has prompted research on the cultivation of *P. palmata* to promote a reliable biomass supply (Martinez et al. 2006; Werner and Diring 2011; Schmedes et al. 2019; Schmedes and Nielsen 2020), reimbursed by the potential application of its derived bioactive hydrolysates (Harnedy et al. 2014; Beaulieu et al. 2016). Like other marine rhodophyte seaweed species, *P. palmata* displays a high nutrient content accommodated by high nutrient uptake capacity for the synthesis of nutrient compounds (Morgan et al. 1980c; Morgan and Simpson 1981a). Hence, several rhodophytes have been suggested as good candidates to enhance nutrient resource utilization when cultivated in water from land-based marine fish farms or adjacent to sea-based farms, a process known as integrated multi-trophic aquaculture or IMTA (Haglund and Pedersen 1992; Chopin et al. 2001; Neori et al. 2004; Sanderson 2006; Abreu et al. 2011; Corey et al. 2013; Grote 2019). The IMTA farming configuration provides high nutrient availability and stimulates the nutrient uptake, which has a positive effect on tissue pigmentation, light-induced stress tolerance, and seaweed productivity (DeBoer and Ryther 1977; Morgan and Simpson

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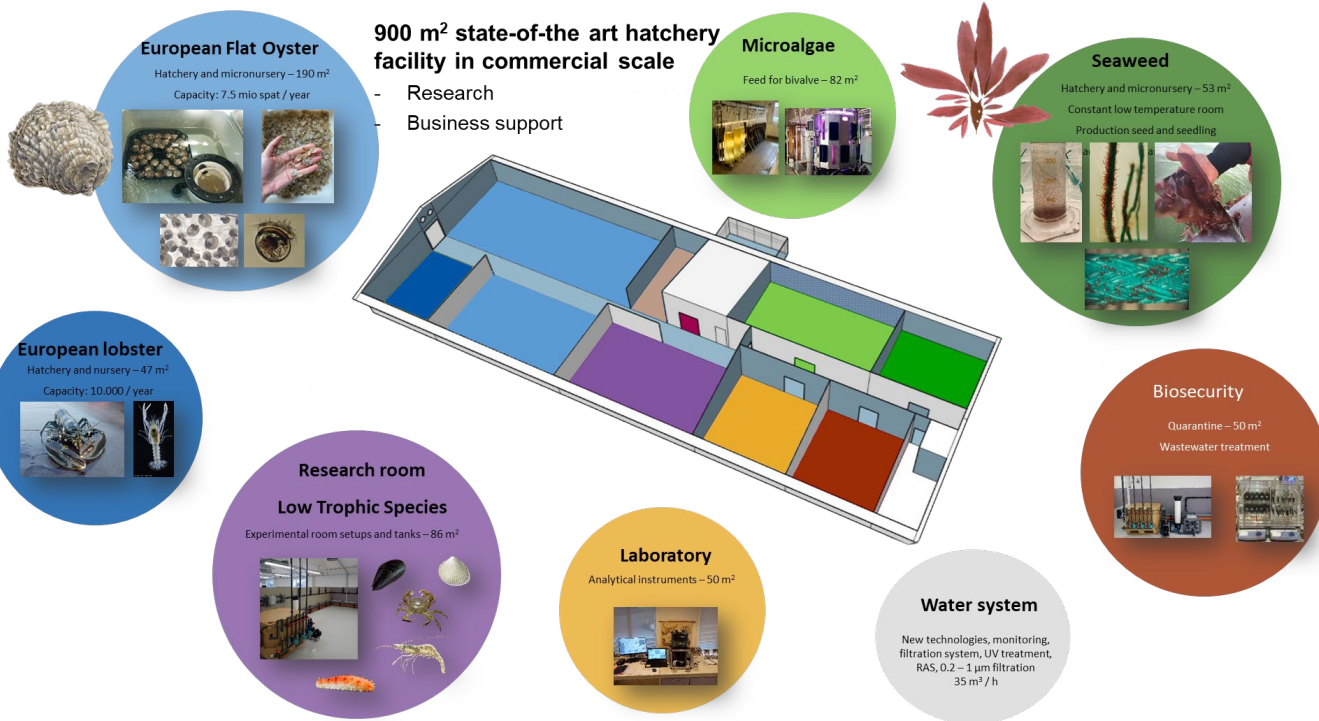
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New facility for increased control

- Research platform
- Business hub



New facility for increased control

- Research platform
- Business hub

FUTURE AMBITIONS

- Diversifying seaweed aquaculture
- Close contact with industry and start-up companies
- Demands for **hatchery development**
 - Innovation and technologies
 - Methodologies and protocols
 - **Species diversification**





Thanks for listening!

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